

## ANTIOXIDANT CAPACITY OF EXTRACTS FROM BULGARIAN MEDICINAL PLANTS

**Tsveta Georgieva\*, Kalin Hristov**

*University of Forestry, Faculty of Veterinary Medicine, Sofia, Bulgaria*

*E-mail: c.georgieva@ltu.bg*

### ABSTRACT

The aim of this research is to establish the redox–modulating capacity of extracts from 5 traditional Bulgarian medicinal plants: *Geranium sanguineum*, *Artemisia annua*, *Tribulus terrestris*, *Cichorium intybus* and *Cotinus coggygia*, comparing their antioxidant capacity at the same concentrations subsequently adding them to the composition of a sperm extender for rams. The methods used include ABTS•+ free radical reduction ability, DPPH• radical reduction, ferric iron reduction by the FRAP method, as well as reduction of cupric ions (Cu<sup>2+</sup>) to cupric ions (Cu<sup>1+</sup>) by the CUPRAC method. As a result, all tested extracts had a high potential for neutralizing free radicals. In conclusion, we can hypothesize that their use also as protective agents in ram sperm diluents will protect gametes from oxidative stress that lowers sperm quality and fertilizing potential.

**Key words:** : DPPH, ABTS•+, FRAP, CUPRAC, antioxidant.

### Introduction

Obtaining ejaculate by artificial vagina, dilution it with sperm extenders, *in vitro* storage of ejaculate at different temperature regimes releases high levels of free radicals, respectively induces high levels of oxidative stress. Depending on activity of the free radicals and their quantity, they can have an extremely adverse effect on the cell (Andreeva *et al.*, 2021). Plasma membrane of ram sperm contains a high percentage of unsaturated fatty acids and a relatively low percentage of cholesterol (Holt, 2000). Examining the level of by–products of sperm membrane lipid oxidation is one of the most widely used methods to assess oxidative stress on gametes (Mojica–Villegas *et al.*, 2014). Numerous studies have shown that the addition of antioxidants to sperm diluents can neutralize free radicals and improve sperm function (Zhang *et al.*, 2012; Agarwal *et al.*, 2014; Gerzilov *et al.*, 2022; Gerzilov *et al.*, 2023). The use of completely synthetic drugs, antibiotics and antimycotics, artificial immunostimulants and chemical growth supplements remains further and further behind (Namkung *et al.* 2004; Oetting *et al.* 2006). The addition of extracts from several Bulgarian herbs will support and even strengthen the action of antioxidants – both enzymatic and non–enzymatic.

The aim of the study is to examine and compare the antioxidant capacity of 5 dry extracts of Bulgarian herbs, dissolved in destilated water at the same initial concentration. According to the results, their antioxidant capacity as protectors of the sperm membrane can be assessed.

### Materials and methods

#### *1. Design of the experiment*

We examined 5 herbal extracts provided by the company Vemo 99, with a certificate for each of them, describing the extraction method and the exact quantitative and qualitative composition of plant metabolites. All herbal extracts are dissolved in distilled water in the same concentration – 1mg/1 ml. For each tested herbal extract, 6 replicates were made. The extracts studied are:

***Tribulus terrestris* L. (Granny's teeth)** – an annual plant of the Zygophyllaceae family. The presence of steroid saponins of the furostanol and spirostanol type and sapogenins has been proven. It also contains flavonol glycosides astragalin, as well as tanning substances and fatty oils.

***Artemisia annua* (Wild Wormwood/Sweet Wormwood)** – all its active ingredients have anti-oxidant and antiviral properties. Artemisinin is the major component and is found in the glandular trichomes of leaves, stems and inflorescences. It is mainly concentrated in the upper parts of the plant, where the new growth also appears. *Artemisia annua* also contains micro- and macroelements such as lead, cadmium and selenium.

***Cotinus coggygia* Scop.** is a tall shrub of the Anacardiaceae family. Contains tannic compounds, determined as tannin, flavonoids, polyphenols determined as catechin, gallotannins, organic acids. Contains also micro- and macroelements – selenium, lead, cadmium.

***Cichorium intybus* L. (Chicory)** belongs to the Asteraceae family and is common in the lowlands of the whole country. The plant is rich in polyphenols, organic acids, mineral acids, inulin and intibin. Coumarins and flavonoids were isolated from the aerial part of the plant, and lactones from the leaves and stems.

***Geranium sanguineum* (Blood Geranium)** is a perennial herbaceous plant of the Geraniaceae family. The composition of the plant includes some flavonoids, catechin tannins, polyphenolic acids, condensed tannins, lactones, anthocyanidis.

## **2. Methods used to determine the antioxidant activity of plant extracts:**

### **Reduction of DPPH• radical**

DPPH is the powdered organic chemical compound 2,2-diphenyl-1-picrylhydrazyl. It is a collection of stable free radical molecules. It is used for antioxidant analysis as well as a standard for the position and intensity of paramagnetic resonance signals. DPPH is a “trap” for other radicals. The decrease in the rate of the chemical reaction upon the addition of DPPH is used as an indicator of the radical nature of the substance under investigation. The reaction is colorimetric and the color change of the reagent is monitored. Measurement of DPPH radical scavenging activity was performed according to the method of Brand-Williams (1995). Reduction of DPPH• to DPPHH results in color changes (from deep violet to light yellow) that are read at 517 nm after 30 min of incubation at room temperature. The percent scavenging activity (AA%) is detected as follows:  $AA\% = 100 - [(Abs(\text{sample}) - Abs(\text{blank}) \times 100) / Abs(\text{control})]$ .

### **ABTS•+ free radical reduction**

The chemical compound ABTS is used to measure the kinetics of specific enzymes. It can be used to indirectly monitor the kinetics of an enzymatic reaction or to quantify the level of hydrogen peroxide. The method is widely used in the food industry. By adding sodium persulfate, ABTS is converted to its radical cation, which is blue in color and absorbs light. The reaction is colorimetric and from the dark blue ABTS radical cation solution, the solution is decolorized. The reaction can be monitored spectrophotometrically. According to the method of Re *et al.* 1999, Trolox is used as a standard and the results are expressed as TEAC % (Trolox equivalent antioxidant capacity vit. E).

### ***Radical reduction of ferric iron by the FRAP method;***

Ferric reduction antioxidant power (FRAP) assay, by the method of Benzie and Strain, 1996 with some modifications. Absorbance was measured at a wavelength of 593 nm.  $\text{FeSO}_4$  is used as a standard and the result is presented as  $\mu\text{M}$  (micromolar)  $\text{FeSO}_4$  equivalent/1g extract. The essence of the reaction is the transition of the iron cation from the third valence to the second valence, and during the reaction, the release of an electron from the reagent and the capture of a free radical leads to a change in the saturation intensity of the solution, it is even possible to discolor it. This method is simple, the result is read colorimetrically and gives sufficient clarity about the antioxidant activity of the investigated substance.

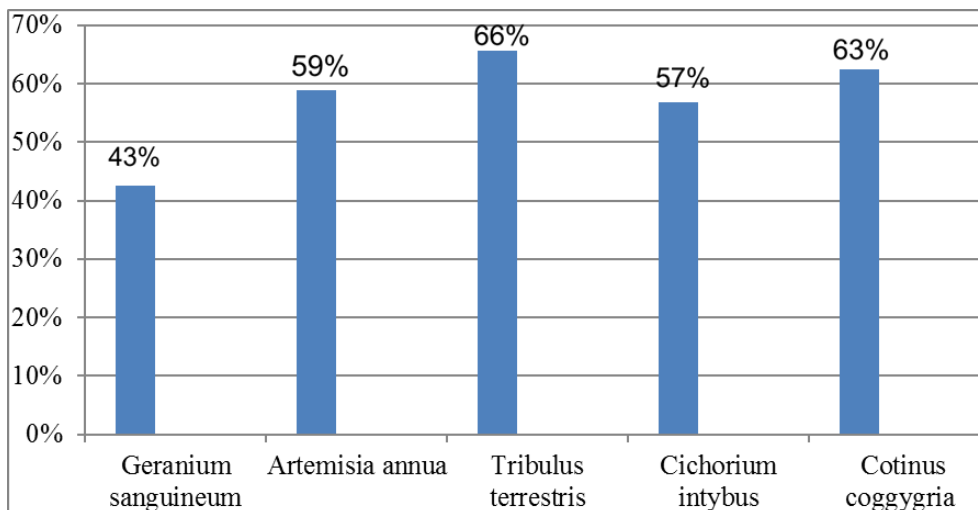
### ***Reduction of cupric ions ( $\text{Cu}^{2+}$ ) to cupric ions ( $\text{Cu}^{1+}$ ) by the CUPRAC method***

Test for reducing antioxidant capacity CUPRAC (CUPric Reducing Antioxidant Capacity) according to the method of Apak *et al.* (2004) with some modifications. The method is based on the reaction of a  $\text{Cu(II)}$ –neocuproine complex (CUPRAC reagent,  $\text{Cu(II)}$ –Nc) with an antioxidant, resulting in a yellow–orange product, a  $\text{Cu(I)}$ –neocuproine chelate complex and an absorption maximum at 450 nm. For the standard curve, Trolox was used at different concentrations ranging from 0.1 mM to 1.0 mM and the results were expressed as CUPRAC [ $\mu\text{M}$  Trolox)/g extract].

## **Results**

All herbal extracts were dissolved in destilated water in concentration 1mg/1 ml and tested by the four methods with the same starting concentration.

When determining the activity by reduction of DPPH• radical method, close to 50% inhibition with the studied concentration was found in *Cichorium intybus* L. ( $56.87 \pm 0.73\%$ ), *Geranium sanguineum* ( $42.56 \pm 1.07\%$ ) and *Artemisia annua* ( $58.85 \pm 0.75$ ). (Figure 1).



**Figure 1: DPPH–capture activity**

The antioxidant activity of the five herbs according to ABTS•+ free radical reduction method. The results are expressed relative to TEAC % (Trolox equivalent antioxidant capacity vit. E). In first place is *Cotinus coggygia* with 6.845%, in second place *Geranium sanguineum* with activity

3.108%, in third place *Cichorium intybus* with activity 0.975 %, in fourth place is *Artemisia annua* – 0.667%, last place is taken by *Tribulus terrestris* – 0.549 % (Figure 2).

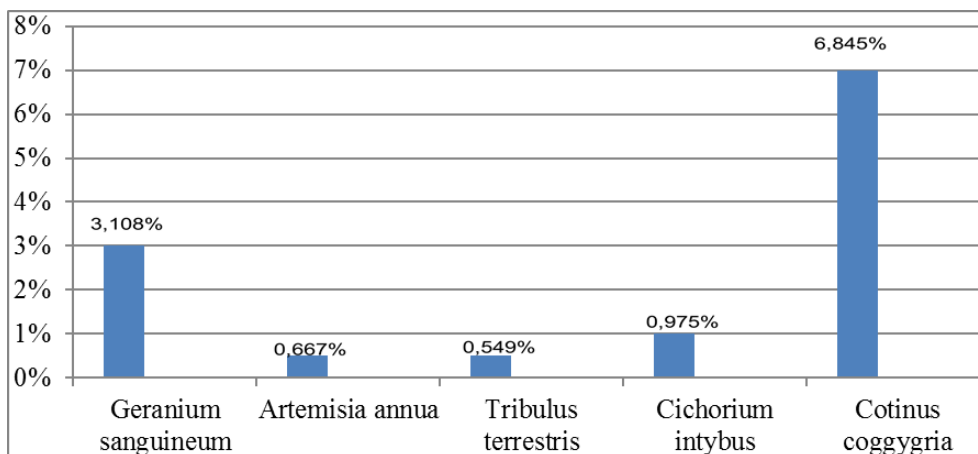


Figure 2: ABTS+ free radical reduction method; TEAC% (Trolox equivalent antioxidant capacity)

The antioxidant activity of the five herbs according to radical reduction of ferric iron by the FRAP method. The results are expressed relative to  $\mu\text{M FeSO}_4$  equivalent/1g extract. In first place is *Cotinus coggygia* with  $51.58 \pm 0.0057 \mu\text{M FeSO}_4$ , in second place *Geranium sanguineum* with activity  $44.96 \pm 0.037 \mu\text{M FeSO}_4$ , in third place *Tribulus terrestris* with activity  $37.08 \pm 0.015 \mu\text{M FeSO}_4$ , in fourth place is *Cichorium intybus* –  $34.37 \pm 0.028 \mu\text{M FeSO}_4$ , last place is taken by *Artemisia annua* –  $31.78 \pm 0.018 \mu\text{M FeSO}_4$  (Figure 3).

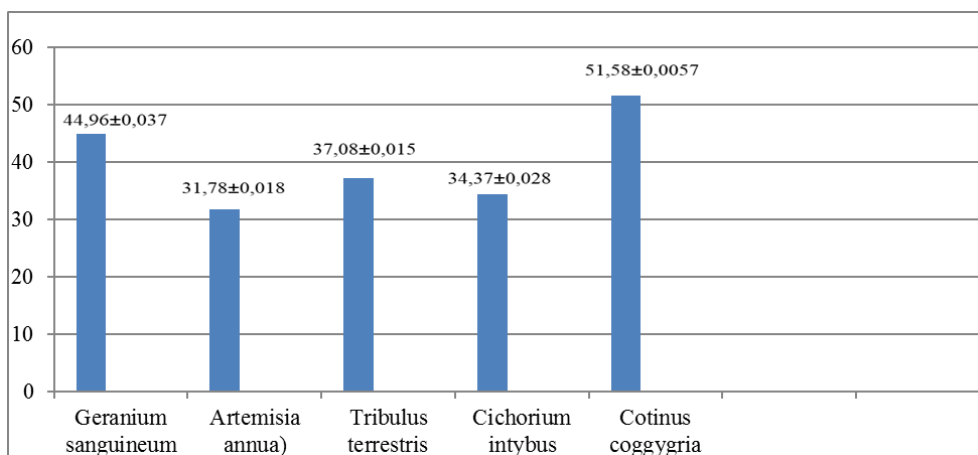


Figure 3: Radical reduction of ferric iron by the FRAP method

The antioxidant activity of the five herbs by reduction of cupric ions ( $\text{Cu}^{2+}$ ) to cupric ions ( $\text{Cu}^{1+}$ ) by the CUPRAC method. The results are expressed relative to TEAC % (Trolox equivalent antioxidant capacity vit. E). In first place is *Cotinus coggygia* with  $1193,565 \pm 0,124 \mu\text{M Trolox equivalent/1mg extract}$ , in second place *Geranium sanguineum* with activity  $1088,83 \pm 0,317 \mu\text{M Trolox equivalent/1mg extract}$ , in third place *Cichorium intybus* with activity  $527,217 \pm 0,111 \mu\text{M}$

Trolox equivalent/1g extract, in fourth place is *Tribulus terrestris*  $425,217 \pm 0,076$   $\mu\text{M}$  Trolox equivalent/1g extract, last place is taken by *Artemisia annua*  $361,87 \pm 0,026$   $\mu\text{M}$  Trolox equivalent/1g extract 9 (Figure 4).

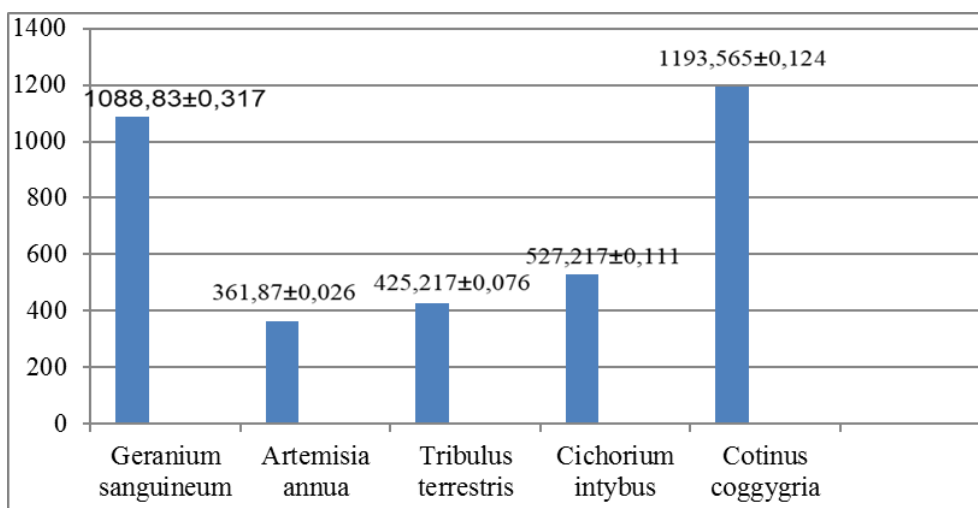


Figure 4: CUPRAC method,  $\mu\text{M}$  Trolox equivalent/1mg extract

## Discussion

There is no common method in the literature that can be used to assess antioxidant capacity (Chochkova *et al.*, 2022). Each method ranks the plant differently, but the trend for highest activity in *Cotinus coggygia* remains. The values of the obtained results give us reason to categorize herbs as plant species with high antioxidant activity and the ability to protect the lipid membrane of spermatozoa from oxidative stress. Free radicals are particles with one or more unpaired electrons that are capable of independent existence (Halliwell and Gutteridge, 2007). The methodology of obtaining ejaculate through an artificial vagina, dilution, storage of sperm at different temperature regimes physical and chemical stress on the sperm membrane which successively reduces the viability of sperm and the ability to fertilization because of the oxidative stress. (Thuwanuta *et al.*, 2011, Andreeva and Stefanov, 2020; Blagova *et al.*, 2021; Andreeva *et al.*, 2023). In the scientific literature, the term free radicals often overlaps and is used synonymously with the term reactive oxygen species (ROS). Of the great variety of ROS, superoxide anion radical ( $\text{O}_2^-$ ), hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) and hydroxyl radical ( $-\text{OH}$ ) are of essential importance for the cell (Andreeva, 2020). In this regard, oxidative stress is considered as the main cause of male infertility, which we can limit by the addition of various antioxidants in the storage media. Our results about the antioxidant capacity of *Cotinus coggygia Scop* coincide with those found by other authors (Georgiev *et al.*, 2014, Kashani *et al.*, 2012, Ngule *et al.*, 2013) The extract contains a high percentage of tannins, gallotannins, flavonoids, organic acids. These substances have high osmoticity and good antioxidant capacity. Furostanol- and spirostanol-type steroidal saponins, sapogenins, the furostanol saponins protodioscin and protogracilin in *Tribulus terrestris* extract are highly active antioxidants (Sun *et al.*, 2002). The leaves also contain the flavonol glycosides astragalin, as well as tannins and fatty oils (Wu *et al.*, 1996; Stefanescu *et al.*, 2020). *Tribulus terrestris* is a widely used antioxidant and stimulant as a food supplement. According to our results the plant has also high local antioxidant capacity. The phenolic

compounds of *Artemisia annua* such as coumarins, flavones, flavonoids, phenolic acids and various phytochemicals – all its active ingredients have antioxidant and antiviral properties. Our results are in agreement with those presented by other authors (Kostova et Dinchev, 2005). The composition of the *Geranium sanguineum* includes some flavonoids, catechin tannins, polyphenolic acids, condensed tannins, lactones, etc. (Leucuta, S *et al.* 2005). Our results about the antioxidant capacity of *Geranium sanguineum* are in unison with those found by other authors (Hardman, *et al.*, 2014; Abbas *et al.*, 2017; Leucuta, 2005). The extract contains a high percentage of tannins, gallotannins, flavonoids, organic acids. *Cichorium intybus*, coumarins and flavonoids are the most active, and lactones are also isolated from the leaves and stems (Al-Snafi, A., 2016). We confirm the research of Al-Snafi *et al.* on the quantitative constituents of the leaves of *Cichorium intybus* because the extract is extracted from this part of the plant.

## Conclusion

All 5 herbs have high antioxidant capacity. Our research leads us to hypothesize that the use of these herbs extracts as protective agents in sperm extenders will protect sperm membrane from oxidative stress, which lowers sperm quality and fertilizing potential.

## Acknowledgements

This work was supported by the project SRS – B 1288/2023 funded by Scientific Research Sector – University of Forestry.

## References

1. Abbas, M., Saeed, F., Anjum, F.M., Afzaal, M., Tufail, T., Bashir, M.S., Ishtiaq, A., Hussain, S., Suleria, H.A.R. (2017). "Natural polyphenols: an overview". *Int. J. Food Prop.*, 20 (8), pp. 1689–1699.
2. Agarwal, A., D. Durairajanayagam, S. S. Du Plessis. (2014a). *Utility of antioxidants during assisted reproductive techniques: An evidence based review* *Reprod. Biol. Endocrinol*, 12: 112
3. Al-Snafi, A. (2016). *Medical importance of Cichorium intybus – A review*. *IOSR Journal Of Pharmacy*, Volume 6, Issue 3 ,PP. 41–56, (e)–ISSN: 2250–3013, (p)–ISSN: 2319–4219
4. Andreeva M. (2020). *Investigation of the effect of breed characteristics insheep on cryotolerance of sperm*. Ph.D. Dissertation. Institute of Biology andImmunology of Reproduction „Akad.Kiril Bratanov”, Bulgarian Academy ofSciences, Sofia, (in Bulgarian)
5. Andreeva M., Alexandrova A., Tsvetanova E., Metodiev N., Stefanov R. (2021). *Effect of the breed on the activity of the antioxidant enzymes – SOD and CAT in ram sperm, before and after cryopreservation*. *Tradition and Modernity in Veterinary Medicine*. vol. 6, No 2(11): 28–33, ISSN 2534–9333, DOI: 10.5281/zenodo.5734021
6. Andreeva M., Metodiev N., Stefanov R., Tsvetanova E., Alexandrova A. (2023). *Evaluation of the cryotolerance of spermatozoa in Sofia (Elin–Pelin) sheep breed*. *Tradition and Modernity in Veterinary Medicine*. 8, 1(14): 11–16. DOI: 10.5281/zenodo.8174279
7. Andreeva M., Stefanov R. (2020). *Influence of the cryopreservation on the vitality of the sperm of the different breeds of rams*. *Tradition and Modernity in Veterinary Medicine*, vol. 5, No 2(9): 26–30. ISSN 2534–9333, DOI: 10.5281/zenodo.4317364
8. Blagova H., Andreeva M., Stefanov R. (2021). *The effect of the degree of dilution on the motility and velocity parameters of sperm in rams*. *Tradition and Modernity in Veterinary Medicine*. vol. 6, No 2(11): 117–120, ISSN 2534–9333, DOI: 10.5281/zenodo.5743820

9. Chochkova M., Georgieva A., Ilieva Ts., Andreeva M., Pramatarov G., Petek N., Petrova P., Štícha M., Mitrev Y., Svete J. (2022). *Hybridization of Aminoadamantanes with Cinnamic Acid Analogues and Elucidation of Their Antioxidant Profile*. Journal of Chemistry, ID 7582587
10. Davies, M.J. (2000). *Electron Paramagnetic Resonance*. Royal Society of Chemistry. p. 178. ISBN 0–85404–310–1.
11. Georgiev, V., Ananga, A., Tsoleva, V. (2014). *Recent advances and uses of grape flavonoids as nutraceuticals*, Nutrients, 6 (1) pp. 391–415
12. Gerzilov V., Alexandrova A., Andreeva M., Tsvetanova E., Georgieva A., Petrov P., Stefanov R. (2022). *Effect of prooxidants and chelator Desferal on the oxidative status and sperm motility of Muscovy semen*. Toxicology Reports, 9, 276–283, DOI: 10.1016/j.toxrep.2022.02.006
13. Gerzilov V., Andreeva M., Tsvetanova E., Georgieva A., Alexandrova A. (2023). *Improvement of diluted semen from Muscovy (Cairina moschata) drakes by addition of water-soluble antioxidants*. Reproduction in Domestic Animals (online). DOI: 10.1111/rda.14487
14. Halliwell, B., J. M. C. Gutteridge. (2007). *Cellular responses to oxidative stress: adaptation damage repair senescence and death*. In *Free Radicals in Biology and Medicine* 4th edn pp. 187–268. Oxford: Oxford University Press
15. Hardman, W.E. (2014). “Diet components can suppress inflammation and reduce cancer risk”. Nutr. Res. Pract., 8 (3), pp. 233–240
16. Holt, W. (2000). *Fundamental aspects of sperm cryobiology: the importance of species and individual differences*. Theriogenology, 53, 47 – 58.
17. Kashani, H., Hoseini, E.S., Nikzad, H., Aarabi, M.H. (2012). *Pharmacological properties of medicinal herbs by focus on secondary metabolites*, Life Sci. J., 9 (1), pp. 509–520
18. Kostova, I., & Dinchev, D. (2005). *Saponins in Tribulus terrestris – Chemistry and Bioactivity*. Phytochemistry Reviews, 4(2–3). <https://doi.org/10.1007/s11101-005-2833-x>
19. Leucuta, S., Vlase, L., Gocan, S., Radu L. & Fodorea C. (2005) *Determination of Phenolic Compounds from Geranium sanguineum by HPLC*, Journal of Liquid Chromatography & Related Technologies, 28:19, 3109–3117.
20. Mojica-Villegas, M. A., J. A. Izquierdo-Vega, G. CharmorroCevallos, M. Sánchez-Gutiérrez. (2014). *Protective effect of resveratrol on biomarkers of oxidative stress induced by iron/ascorbate in mouse spermatozoa*. Nutrients, 6: 489 – 503. (doi:10.3390/nu6020489)
21. Namkung, H., Li J. Gong, M., Yu, H., Cottrill, M., & de Lange, C. F. M. (2004). *Impact of feeding blends of organic acids and herbal extracts on growth performance, gut microbiota and digestive function in newly weaned pigs*. Canadian Journal of Animal Science, 84(4).
22. Ngule, C.M., Anthoney, S.T., Obey, J.K. (2013). *Phytochemical and bioactivity evaluation of senna didymobotrya fresen irwin used by the nandi community in Kenya*, Int. J. Bioassays, 2 (7), pp. 1037–1043
23. Oetting, L. L., Utiyama, C. E., Giani, P. A., Ruiz, U. dos S., & Miyada, V. S. (2006). *Efeitos de extratos vegetais e antimicrobianos sobre a digestibilidade aparente, o desempenho, a morfometria dos órgãos e a histologia intestinal de leitões recém-desmamados*. Revista Brasileira de Zootecnia, 35(4).
24. Sharma, Om P.; Bhat, Tej K. (2009). *"DPPH antioxidant assay revisited"*. Journal of Food Chemistry. 113 (4): 1202–1205. doi:10.1016/j.foodchem.2008.08.008
25. Sun, W., Gao, J., Tu, G., Guo, Z., & Zhang, Y. (2002). *A New Steroidal Saponin from Tribulus Terrestris Linn*. Natural Product Letters, 16(4). <https://doi.org/10.1080/1057563029002054>

- 
26. Thuwanuta, P., K. Chatdarong, A. S. Bergqvist, L. Söderquist, K. Thiangtum, D. Tongthainan, E. Axné. (2011). *The effects of antioxidants on semen traits and in vitro fertilizing ability of sperm from flat-headed cat (Prionailurus planiceps)*. Theriogenology, 76: 115 – 125
  27. Wu, G., Jiang, S., Jiang, F., Zhu, D., Wu, H., & Jiang, S. (1996). *Steroidal glycosides from Tribulus terrestris*. Phytochemistry, 42(6). [https://doi.org/10.1016/0031-9422\(96\)00182-3](https://doi.org/10.1016/0031-9422(96)00182-3)
  28. Zhang, W., K. Yi, C. Chen, X. Hou, X. Zhou. (2012). *Application of antioxidants and centrifugation for cryopreservation of boar spermatozoa*. Anim. Reprod. Sci., 132: 123–128
  29. Zhen, J.; Villani, T. S.; Guo, Y.; Qi, Y.; Chin, K.; Pan, M. H.; Wu, Q. (2016). *Phytochemistry, antioxidant capacity, total phenolic content and anti-inflammatory activity of Hibiscus sabdariffa leaves*. Food Chemistry. 190: 673–680. doi:10.1016/j.foodchem.2015.06.006. PMID 26213025.
  30. Zhong, Y, F. (2015). Shahidi, in *Handbook of Antioxidants for Food Preservation*. „Ferric Reducing Antioxidant Power Assay“ 12.3.2.1